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EFFECT OF PLANT EXTRACTS ON RADIAL GROWTH OF *Helminthosporium oryzae* CAUSATIVE OF BROWN SPOT DISEASE OF RICE UNDER *IN-VITRO*

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ABSTRACT

Brown spot disease, caused by *Helminthosporium oryzae*, is worldwide problem capable of causing considerable damage to paddy in the nursery, field or grain yield. The disease is seed borne, and thus can be transmitted through infected seeds and crop residues, alternate hosts and contaminated irrigation water. The objective of this study was to evaluate the effect of plant extracts on radial growth of *Helminthosporium oryzae* on rice plants. An *in-vitro* experiment was conducted at the Plant Pathology Laboratory of National Root Crop Research Institute, Umudike, Abia State, in Nigeria. Treatments included water and alcohol extracts of *Azardiractha indica* (Neem leaves), *Piper guineensis* (seeds), *Garcinia cola* (Bitter cola seeds), *Ocimum gratissimum* (leaf) and *Vernonia amygdalina* (leaf); and synthetic fungicide (Benomyl) at a concentrations of 10, 25 and 30% of the extract applied to *H. oryzae* in culture. The test materials were administered on *Helminthosporium oryzae*, sourced from rice seeds and infected shoot system of rice. Alcohol extract of *Piper guineensis* had the highest radial growth inhibition (89.89%) by the fifth day, but was not significantly different from *Azardiractha indica*, which had an inhibition value of 81.02%. The least effective plant extract was *Ocimum gratissimum* with radial inhibition of 11.50%, which occurred also on the fifth day. Plant extracts were as effective as the synthetic fungicide in inhibiting growth of the test fungus. Therefore, the effective extracts, all of which are readily available to the farmers, should be promoted instead of the synthetic fungicides, which are in limited supply and invariably expensive for rice farmers in Nigeria.

Key Words: *Helminthosporium oryzae*, *Piper guineensis*, *Zinzigiber officinale*

RÉSUMÉ

La maladie des taches brunes, causée par *Helminthosporium oryzae*, est un problème mondial susceptible de causer des dommages considérables au riz en pépinière, champ et en rendement en grains. La maladie est transmise par les semences et peut donc être transmise par des semences et des résidus de culture infectés, des hôtes alternatifs et de l'eau d'irrigation contaminée. L'objectif de cette étude était d'évaluer l'effet d'extraits de plantes sur la croissance radiale d'*Helminthosporium oryzae* sur les plants de riz. Une expérience *in vitro* a été menée au laboratoire de phytopathologie de l'Institut national de recherche sur les cultures racines, Umudike, État d'Abia, au Nigéria. Les traitements comprenaient des extraits aqueux et alcooliques d'*Azardiractha indica* (feuilles de Neem), de *Piper guineensis* (graines), de *Garcinia cola* (graines de cola amer), d'*Ocimum gratissimum* (feuille) et de *Vernonia amygdalina* (feuille); et fongicide synthétique (Benomyl) à des concentrations de 10, 25 et 30% de l'extrait appliqué sur *H. oryzae* en culture. Les matériaux d'essai ont été administrés sur *Helminthosporium oryzae*, provenant de graines de riz et d'un système de pousses de riz infecté. L'extrait alcoolique de *Piper guineensis* avait l'inhibition de la croissance radiale la plus élevée (89,89%) au cinquième jour, mais n'était pas significativement différente de celle d'*Azardiractha indica*, qui avait une valeur d'inhibition de 81,02%. L'extrait de plante le moins efficace était *Ocimum gratissimum* avec une inhibition radiale de 11,50%, qui s'est également produite le cinquième jour. Les extraits végétaux étaient aussi efficaces que le fongicide synthétique pour inhiber la croissance du champignon d'essai. Par conséquent, les extraits efficaces, qui sont tous facilement disponibles pour les agriculteurs, devraient être encouragés à la place des fongicides synthétiques, qui sont en quantité limitée et invariablement coûteux pour les riziculteurs au Nigeria.

Mots Clés: *Helminthosporium oryzae*, *Piper guineensis*, *Zinzigiber officinale*

INTRODUCTION

The key disease to viable rice production in Nigeria is Brown spot disease, caused by *Helminthosporium oryzae* (Grist, 1975; Imolehin, 1983; Ekeleme *et al.*; 2008). It is distributed world wide and causes considerable damage to paddy rice in nurseries, field and shrinkage of grain yield. The disease is seed borne and is thus transmitted to healthy crops through infected seeds and crop residues, alternate hosts and contaminated irrigation water (Samson and Kadiri, 2007).

Unlike other fungal diseases of rice, Brown leaf spot is more severe on older plants than on seedlings; and is characterised by dark brown oval spots (about 3.8 cm long) on the leaves (Grist, 1975). The larger spots usually have lighter coloured centres and dark brown margins. Cultural control measures include burning all crop residues and alternative hosts and use of seeds from healthy plants (Aquino *et al.*, 1985).

Application of chemicals such as Seed-plus, Dress force, Apron star or Super Homai at the rate of 10 g per 5 kg of seeds before sowing has been effective in controlling this seed borne pathogens (Groth and Hollier, 2000, NCRI, 2008). When symptoms are observed, Ekeleme *et al.* (2008) recommended a spray of Dithane M-45 (Mancozeb) weekly at the rate of 1.5 kg ha⁻¹ (about 2-3 small match boxes full per 15 L sprayer) for 3 weeks as an effective control measure. The objective of this study was to evaluate the effect of plant extracts on radial growth inhibition of *Helminthosporium oryzae* on rice plants as an alternative to synthetic fungicides.

MATERIALS AND METHODS

Preparation of plant extracts. Extract were prepared from fresh leaves of *Azardiractha indica* (Neem leaves), *Piper guineensis* (seeds), *Garcinia cola* (Bitter cola seeds), *Ocimum gratissimum* (leaf) and *Vernonia amygdalina*

(leaf), using sterile distilled water and absolute alcohol of 98% purity. The leaves or seeds in their respective cases, were thoroughly washed in running tap water, and subsequently with distilled water, before being air dried at 27 °C for twelve hours. The materials, which were not completely dried were smashed using a warring blender to obtain approximately 1 kg of paste of each test material.

Water extraction. Water extracts were obtained by adding 10, 25 and 30 g of each paste to 100 ml of sterile distilled water in 250 ml beakers; and stirring vigorously before allowing to stand for 1 hour. Each plant material was extracted separately, with intermittent cleaning of the warring blender using sterile distilled water. The supernatant liquid of each extract was passed through a Whatman No 1 filter paper. The filtrate was also passed through a membrane filter (0.22 µm) to avoid bacterial or fungal contamination. The resulting filtrate was later concentrated by autoclaving and allowed to cool to room temperature, before being stored under aseptic refrigerator conditions, in properly corked and labelled Mac Carthney bottles. The interface between the corks and the bottles were sealed with masking tapes, to prevent contamination.

Alcohol extraction. For alcohol extraction, each plant material was oven dried at 45 °C in for 24 hours after being washed three times with distilled sterile water, and air-dried at room temperature. Each plant material was ground with a warring blender to obtain approximately 1 kg powder each. Each powder of 10, 25 and 30 g was extracted with 95% ethanol (100 ml) and concentrated through a rotary vacuum pump flash evaporator, to syrup form weighing 26 g from each powder. The syrup residue obtained was diluted to 2.5 g l⁻¹. Preparations were made in triplicates throughout the experiment.

Growth of *Helminthosporium oryzae*. One disc (3 mm diameter) of an 8 day-old culture of each fungus was placed in each of three

petri dishes (9 cm diameter) containing 20 ml of potato dextrose agar (PDA) medium and 3 ml of leaf extract (Amadioha, 2003). Three concentrations of the extracts were tested, namely 10, 25 and 30 g of each powder to 100 ml of absolute (98%) alcohol and 0 (sterile water) as control. Each of these concentrations was replicated three times. These were tested on the radial growth of three fungal pathogens, namely, *Fusarium moniliforme*, *Helminthosporium spp* and *Phoma spp*; each fungus was tested separately. Sterile distilled water was used as the control. Twelve Petri dishes were used for each plant extract, against one fungus at a time. Another twelve dishes were used for the synthetic fungicides at concentrations of 10, 25 and 30 g per 100 ml of distilled water. Each concentration was replicated three time with distilled water as control; and laid out in a randomised complete design (CRD). The whole set up of Petri dishes was incubated at 27 °C, with 12 hours of alternating light and darkness.

Determination of radial growth. Radial growth of the test fungi was measured daily for a period of seven days starting from the second day after inoculation. The diameter of the fungal colony was measured from the back sides of the plates, using a metre rule. Colony growth was measured by taking the average of the largest and shortest diameter of the same colony per plate. This was because colony growth of fungi was not always in regular circles. The fungitoxicity of the extracts was calculated in terms of percentage colony inhibition using the formula of Amadioha (2003):

$$\text{Growth inhibition (\%)} = \frac{dc - dt}{dc} \times \frac{100}{1}$$

Where:

dc is the average diameter of fungal colony with control; and dt is the average diameter of fungal colony with treatment.

Data analysis. Data collected were subjected to Analysis of Variance (ANOVA) using GenStat 7.2 DE version (2007). Mean separation was done using Fisher's Least Significant Difference (F-LSD) at 5% level of probability.

RESULTS

Radial growth of *Helminthosporium oryzae*.

There were significant negative radial growth effects of plant extracts of both aqueous and alcohol extraction, on radial growth of *Helminthosporium oryzae* in culture (Table 1). *Zingiber officinale* extracts in alcohol had the highest radial growth inhibition effect (94.96%), but was not significantly different from the effect of *Azadirachta indica*, which had inhibition value of 92.07%. The lowest growth inhibition was in *O.gratssimum*, by aqueous extraction method, which had radial inhibition of 16.50% right on the first day. On the second day, alcohol extract of *Piper guinensis* had the highest (86.30%) inhibition value; followed by alcohol extract of *Z. officinale* (82.30%). The least effect was in aqueous of *G. cola* with inhibition value of 16.60%.

On the third day, alcohol extract of *Piper guinensis* had the highest inhibition value of 89.89%; followed by alcohol extract of *Azadirachta indica* (81.02%). The least effect in this case was in aqueous extract of *O. gratssimum* with value of 11.50%. The five plant extracts by both water and alcohol extractions performed better than the control.

Interaction effects of plant extracts x concentrations.

For Bitter cola, there was significant interaction effect among the three concentrations x plant extract combinations (Table 1). In alcohol, the highest reduction was 92.52% obtained in 30 g of the extract on the first day. This was followed by 86.70% achieved with 30 g of the extract by alcohol extraction on the third day. The lowest values were obtained in water extraction.

For Neem, the interaction effect of plant extract x 10 g by alcohol extraction, gave comparatively the highest radial inhibition on third (90.60%) and fifth (85.41%) days in culture, compared to plant extract x 25 g and plant extract x 30 g interaction effects. It was also observed in Neem that the inhibition effect depreciated with time as higher radial inhibitions were observed in the first day (95.40%) than in subsequent days (Table 1). This trend was observed in *Z. officinale*, but was different in *O. gratssimum* and *P. guinensis*, where positive interaction effects were observed with increase in concentration of plant extracts.

Compared to the standard fungicide (Benlate), it was clear that phytochemical effects due to alcohol extraction performed better for all the plant materials used; except for *O.gratssimum* in which radial growth was inhibited more by the action of the synthetic fungicide.

DISCUSSION

Effects of plant extracts on radial growth of *Helminthosporium oryzae* in culture.

The study showed that the various plant extracts and the synthetic fungicide inhibited the radial growth of *Helminthosporium oryzae* in culture. *Piper guineesis* was found to perform best in suppressing mycelial growth of *H. oryzae*; followed by *Azadirachta indica*; while the least was *Occimum gratissimum*. The significant differential effect of the various plant materials could be as a result of the different bioactive components of each botanic extract. According to Amadioha (2003), the active agent in *A. indica* is *Azadiratin*; while those in *Z. officinale* consist of linalole, imonene, Zingerene and Linoelic acid. In *Garcinia cola*, the active ingredients responsible for anti-microbial, anti-viral and anti-inflammatory properties are bioflavonoids, xanthenes and benzophenomes (Kikuzaki and Nakatani, 1993; Derbalah, 2011). According to them, some of these compounds like polyphenols have been

TABLE 1. Percentage growth inhibition of *Helminthosporium oryzae* by the treatments in first, third and fifth days

Treatment	Incubation period (days), Concentration (%) and growth inhibition (%)														
	1 day					3 days					5 days				
	0	10	25	30	Mean	0	10	25	30	Mean	0	10	25	30	Mean
<i>Garcinia cola</i>															
Water extract (WE)	0	35.70	38.10	34.00	27.00	0	15.40	24.70	26.20	16.60	0	28.90	29.60	33.50	23.00
Alcohol extract (AE)	0	53.27	39.26	92.52	71.51	0	28.20	36.50	86.70	63.20	0	43.84	26.29	75.39	61.62
<i>Azardiractha indica</i>															
Water extract	0	61.50	68.90	68.90	49.80	0	61.40	69.10	72.70	50.80	0	68.00	58.10	83.10	52.30
Alcohol extract	0	95.40	95.41	76.47	92.07	0	90.60	68.80	60.75	67.31	0	85.41	71.00	66.18	81.02
Zingiber officinale															
Water extract	0	44.90	25.20	34.20	26.10	0	31.00	25.30	32.00	22.10	0	27.10	18.50	23.60	17.30
Alcohol extract	0	95.09	95.09	88.66	94.96	0	86.20	86.20	56.90	82.30	0	47.50	57.65	55.23	65.33
<i>Ocimum gratissimum</i>															
Water extract	0	21.60	21.40	23.00	16.50	0	31.00	22.30	25.40	19.70	0	2.20	25.50	18.30	11.50
Alcohol extract	0	19.47	13.79	94.83	57.02	0	36.10	68.30	76.30	70.20	0	24.79	63.18	69.42	64.72
Piper guinensis															
Water extract	0	33.30	22.20	33.30	22.20	0	24.30	33.10	33.70	22.80	0	16.20	20.50	21.70	14.60
Alcohol extract	0	70.35	95.15	95.15	90.42	0	67.90	89.30	88.10	86.30	0	72.95	93.24	91.88	89.89
Benlate	0	65.00	62.80	56.10	46.00	0	57.40	67.30	66.20	47.70	0	34.80	51.70	51.90	34.60
LSD 0.05 (WE)		1.39			0.69		1.57			0.78		2.41			1.71
LSD 0.05 (AE & Benlate)		17.95			8.98		23.90			11.95		22.85			11.42

WE = Water Extraction and AE = Alcohol Extraction.

shown to express their antifungal action through membrane perturbations. This disruption of the cell membrane, coupled with the actions of lysolytic compounds on the transpeptidation of the cell membrane, could lead to enhanced antimicrobial effect of the biocompounds. Opara and Agugo (2014) also observed that hot water extract of *P. guinensis* had better performance than *Z. officinale* in inhibiting growth of bacteria that caused post-harvest loss of tomato in Abia State, Nigeria.

Effect of concentrations of bioactive compounds of the plant extracts on radial growth of *Helminthosporium oryzae* in culture. The fungitoxicity of the plant extracts against mycelial growth increased as the concentration increased in the plant extracts. This is in agreement with Bailen *et al.* (2013), who reported that the mortality of *B. brassicae* using plant extracts, increased with increasing concentrations and exposure periods. According to them, the physical and chemical properties of the essential oils exhibited different persistent levels of pesticide properties and different action mechanisms. Suleiman and Taiga (2009) reported that the crude extract of *G. cola* fruits showed significant reduction in mycelial growth of *Pythium aphanidermatum* at different levels of concentrations, which is in agreement with the result of this study. According to them, high fungi toxicity *in vitro* was observed at 80 and 100% concentrations.

Effect of differences in extracting solvents on inhibitory activities of the plants bioactive compounds. Alcohol extracts had higher inhibition effects than the aqueous counterparts (Table 1). This could be as a result of higher solvent and extractability of alcohol of phytochemical substances used in the study, compared to water extraction. This was observed in all the plant phytochemicals used. This is in agreement with the findings of Ngo *et al.* (2017) in a study on the 'impact of different extraction solvents on bioactive

compounds and antioxidant capacity from the root of *Salacia chinensis* L.', where they observed that absolute methanol had the highest extractable solids (15.6%), followed by 50% ethanol, 50% methanol, and 50% acetone (14.3%, 12.3%, and 12.2%, resp.). They also reported that water extracted half of extractable solids in comparison with absolute methanol.

Interaction effects of plant extracts and concentration of bioactive phytochemicals.

Interaction effects of plant bioactive compounds and concentration showed that there was significant inhibitory effects on radial growth of the test fungus (Table 1). It was also observed in *O. gratissimum* by alcohol extraction that the inhibition effect appreciated with time as lower mean radial inhibitions were observed in the first day (19.47%) than in subsequent days. This could mean that the actions of the bioactive compounds of the various botanicals were more effective with concentration and time of exposure on the target organism. This is consonance with the findings of Ahmed *et al.* (2020), who in their study 'insecticidal activity and biochemical composition of *Citrullus colocynthis*, *Cannabis indica* and *Artemisia argyi* extracts against cabbage aphid (*Brevicoryne brassicae* L.)', reported that the percent mortality of *B. brassicae* to be directly related to the concentration of the plant extracts and the exposure period. According to them, maximum mortality was recorded after 72 hr of exposure to the *A. argyi* extract at a concentration of 20 mg mL⁻¹ and caused 88.33±3.87% mortality; while after 48 hr of exposure, the mortality was recorded as 60.00± 2.27%. Depreciation in inhibitory effects of the active ingredients was noticed in Neem by alcohol extraction. This might have been caused by negative effects of some environmental factors such as heat which might have led to increased breakdown of the fungitoxic substances in the plant extracts. Also, the decrease in fungitoxic effect of the

bioactive substance with time experienced in Neem plant extract might have been as a result of reaction of these active substances with some atmospheric chemical elements, thereby resulting to less fungitoxic compounds.

The oxidation of phenolic compounds using oxygen and formic acid as indirect oxidants (Javier, *et al.*, 2014). According to them, this novel immobilised tri-enzyme system removes important pollutants such as hydroxylated aromatic derivatives (phenol, 4-aminophenol, 2,4-dichloro-phenol or α -naphthol) using formic acid and molecular oxygen as substrates. In addition, this system generates CO₂ as waste beyond the oxidised phenols that can be easily separated from the aqueous solution. Also, Alnaizy and Akgerman (2000) reported in their experiment 'Advanced oxidation of Phenolic compounds' that Phenol degradation with a UV/H₂O₂ advanced oxidation process gave the reaction products which included hydroquinones, benzoquinones, and aliphatic carboxylic acids with up to six carbon atoms which do not have the same bioactivity with those that were not oxidised.

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